Genotyping of Streptococcuspneumoniae Isolated from Upper Respiratory Tract Infections by RAPD analysis

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Streptococcus pneumoniae, commonly called the pneumococcus, is responsible for high rates of morbidity and mortality worldwide. Genotyping of S. pneumoniaeby using RAPD technique have largely replaced the phenotyping methods. this study was suggested and designed to study the genotyping of S. pneumoniaeby using RAPD analysis method. In this study, randomly amplified polymorphic DNA (RAPD) analysis was performed by using arbitrarily primer (S1254) and PCR technique. Strains were considered different from one another if their patterns differed by one prominent band in three repeated experiments.

In the current study, 31 patterns (different types of S. pneumoniae) were yielded out of 32 isolates composed 4-10 fragments ranging from (64-1922) pb in size of which one permenant band (134 pb) was obtained from most isolates. The dendrogram generated when these (32) different isolates of S. pneumoniae were compared is and this dendrogram appeared two main clusters, whereas the first includes 26 isolates with 3 subcluster, and the second include the remaining 6 isolates with two subcluster. The first contain 4 isolates and the second included 2 isolates. The higher homology 66% were appeared between isolates S_{13} and S_{15} , followed by 60% between isolates S_{1} and S_{32} . These of RAPD-PCR method it possible to identify non-serotyped strains, and shows the necessity of this molecular typing technique for typing S. pneumoniaestrain from patients with LRTI.

Intruduction:

S.pneumoniae is the major cause of community- acquired pneumonia in adults and serious respiratory infections in children in the United States. An estimated three to five million deaths occur annually in children under 5 years of age due to acute respiratory infections, for which Streptococcus pneumoniais the most important pathogen (Obaro, 2011). Differentiation of S. pneumoniafrom other viridans group streptococci, has conventionally been based on phenotypic characteristics, most commonly by demonstrating optochin OPT susceptibility and/or solubility in bile (Facklam, 2002).

Molecular genetic analyses based on the 16S rRNA gene have provided a potent means of characterization at the species level (Stackebrandt and Goebel, 1994). Unfortunately, S. pneumoniae shares over 99% 16S rRNA gene identity with S. mitis and S. oralis (Suzuki etal., 2005), complicating molecular identification among them. Genetic transformation, exchanges of virulence factors among streptococcal species and the appearance of new closely related streptococcal species, such as S. pseudopneumoniae, confirm that problems in taxonomy and pneumococcal identification will continue in the future (Arbique et al., 2004; Fink, 2005).

Molecularmethods for typing are useful in studying a short-term (the spread of an isolate of a hospital or local community) as well as epidemiological follow-up. For S.pneumoniaeepidemiology, accurate typing methods, such as pulsed-field gel electrophores (PFGE) and restriction fragment length polymorfism (RFLP) (Schlegel et al., 2003) and multilocus sequence typing (MLST) (Spratt 1999), have been developed. The PFGE method is based on the evaluation of total chromosomal DNA, and the obtained PFGE type is compared with similarity and diversity patterns. It is commonly used and considered as a gold standard (Shaaly et al., 2005). Several modifications of the RFLP analysis have been used in the epidemiological follow-up of S.pneumoniae(Doit et al., 2002; Schlegel et al., 2003).

Molecular studies on S. pneumoniae have received a little attention in Iraq. In light of the medical importance and to demonstrate the molecular relation between isolates, this study was suggested and designed to study the Genotyping of S. pneumoniae by using RAPD analysis method.

Materials and Methods

Bacterial strains.

In this study 74 pneumococcal isolates recovered from 600 sputum of patients with clinical symptoms of LRTI (pneumonia, COPD and TB)During theperiod of from November, 2012 - February, 2013 obtained from consultants clinic for respiratory and chest in Al-Sadder Medical City .All isolates were diagnosis by microscopic ,colonial morphology,and by used Conventional biochemical test . STREPO-SYSTEM 9R and VITEK-2 copact using GPcards were used for confirmed and accurte final diagnosis test ,

DNA extraction.

For genetic analysis, we selected 32 of S.pneumoniae isolates . S.pneumoneae isolates were cultured on tryptic soy agar suplimented with 5% sheep blood and inoculated individually into TSB and incubated at 37°C/24h. Genomic DNA Extraction Kit (Geneaid) was used for DNA extraction.

Detection of DNA by Agarose Gel Electrophoresis

Gel electrophoresis was used for detection of DNA by UV transilluminator according to Sambrooket al. (2001).

Genotyping of Isolates by RAPD Method

An arbitrary primer was used for RAPD-PCR assay of S.pneumoniae DNA sample (100 ng) using a modified protocol of Duarte etal., (2005) in a 25- μ l PCR reaction volume. And RAPD-PCR products were resolved by electrophoresis on ethidium bromide (0.5 μ g/ml) pre-stained 1.0% agarose gel using a 1 Kb DNA ladder (Fermentas) for size extrapolation. The bands were detected by used UVI B and software (version 12.14). Isolates were considered different from one another if their patterns deferred by one prominent band in three repeated experiments.

| Targete gene | DNA sequence (5'-3') | Reference |
|--------------|----------------------|----------------------|
| RAPD1254 | CCG CAG CCA A | Duarte et al. (2005) |
| | | |

Table (1): Program used to amplify the RAPD 1254

| Stage | Temperature (time) | |
|----------------------|--------------------|---------|
| Initial denaturation | 94C for 5min | 2cycle |
| Denaturation | 94C° for 1min | |
| Annealing | 37C° for 1min | 40cycle |
| Extension | 72C° for 2min | |
| Final extension | 72C° for 10min | |

Results and Discussion:

In this study, randomly amplified polymorphic DNA (RAPD) analysis was performed by using arbitrarily primer (S1254) that described by Duarte et al. (2005) and PCR technique. The PCR-RAPD products were resolved by 2% agarose gel electrophoresis. Strains were considered different from one another if their patterns differed by one prominent band in three repeated experiments.

In the current study, 31 patterns (different types of S. pneumoniae) were yielded out of 32 isolates composed 4-10 fragments ranging from (64-1922) pb in size of which one permenant band (134 pb) was obtained from most isolates (figure 1).



Figure (1) RAPD amplified products of the S. pneumoniaeusing random primers (1254)

These of RAPD-PCR method it possible to identify non-serotyped strains, and shows the necessity of this molecular typing technique for typing S. pneumoniaestrain from patients with LRTI.

The dendrogram generated when these (32) different isolates of S. pneumoniaewere compared is shown in figure (2). This dendrogram appeared two main clusters, whereas the first includes 26 isolates with 3 subcluster, and the second include the remaining 6 isolates with two subcluster. The first contain 4 isolates and the second included 2 isolates (figure 2). The higher homology 66% were appeared between isolates S_{13} and S_{15} , followed by 60% between isolates S_1 and S_{32} .

Kilian et al., (2008) investigated the evolutionary history of the pneumococcus and its close commensal relatives using a polyphasic phylogenetic strategy and analysed of this unique cluster of closely related species with very distinct pathogenic potentials. Surprisingly, Arbiqueet al., (2004) demonstrated that S. pneumoniae is but one of several hundred distinct phylogenetic lineages of a cluster of otherwise

commensal streptococci known as S. mitis or S. pseudopneumoniae in which S. pneumoniae is no more genetically divergent from other members of the cluster than individual lineages of S. mitis are from each other. The name S. pseudopneumoniae was recently assigned to one of these lineages.

Because the number of strains has been increasing year by year, the possibility of an alternative method for typing pneumococci was investigated. Therefore, a number of genotyping techniques such ribotyping (Shundi et al., 2000), pulse field gel electrophoresis (Gonzalez–Rey et al., 2003), random amplified polymorphic DNA (RAPD) analysis (Fica et al., 2003) and multilocus sequence typing (Enright et al., 2001) are being applied worldwide for typing of isolates. In order to simplify the typing work and reduce cost and time, the Fenollet al., (1997) used RAPD test routinely in the laboratory, based on the SGT distribution of the pneumococci isolated in Spain.

RAPD have been also used for subtyping of GAS isolates mainly in western countries (Gonzalez–Rey et al., 2003; Fica et al., 2003). The RAPD assays were performed at constant DNA concentrations and the experiments were repeated several times by taking proper precautions. In the present study, RAPD results were reproducible when samples were run in large gels simultaneously. Thus, all these facts suggest the usefulness of this technique in disease outbreak detection. Nandi et al., (2008) demonstrated the benefit of RAPD fingerprinting in comparison to other molecular methods in identifying and characterizing S. pneumoniae isolates obtained from pharyngitis and RF/RHD cases.

In addition use of PCR based RAPD method for typing of GAS was found to be highly discriminatory. As reported earlier, selection of primers, optimization of PCR condition and combination of different primers play an important role in discriminating the isolates by RAPD (Seppälä et al., 1994). Nandi et al., (2008) used five arbitrarily selected primers were tested. The majority of arbitrary primers used, produced distinctly reproducible patterns in all the isolates studied. These result confirm the result of the present study in which the primer used (S1254) in this study gave a high difference between isolates. Although, Iwalokun et al., (2012) demonstrated the limitations of RAPD in epidemiological characterization of S. pneumoniaeisolates in this environment. Therefore, for better phylogenetic grouping of S. pneumoniaeand improved understanding of serotype switching, typing techniques such as multilocus sequence typing (MLST) are required and this method was used for better understanding of clonal diversity, dissemination and pathogenicity of S. pneumoniaeat regional, national and country levels.

Genotypic similarity, as was found in the majority of the paired samples, indicates clonal relatedness and implicates that pneumococci present at different sites in the upper respiratory region of a patient originate from a single source (Tonnaeret al., 2005). In conclusion the primer used in this study is the best for genotyping of S. pneumoniaedue to it gives high varieties.

Despite advances in our understanding of the epidemiology and pathogenesis of pneumococcal infections, the precise genetic factors that predispose a given pneumococcal clone for disease versus carriage remain unknown (Pettigrew et al., 2006).

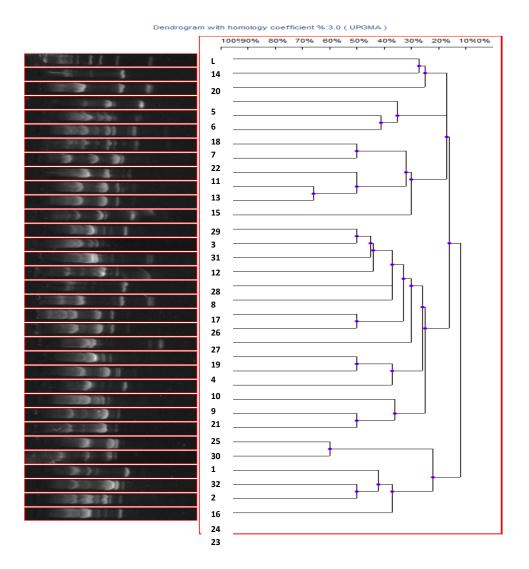


Figure (2): Dendrogram showing genetic relationship of 32 S. pneumoniae isolates produced by RAPD with primer 1254.

Conclusion:RAPD analysis is the faster and simplest method used for typing of S. pneumonia by using primer S1234.

References

- **Arbique**, J.C.; Poyart, C.; Trieu-Cuot, P.; Quesne, G.; Carvalho, G.; Steigerwalt, A.G. and Morey, R.E. (2004). Accuracy of phenotypic and genotypic testing for identification of Streptococcuspneumoniae and description of Streptococcuspseudopneumoniae sp. nov. J. Clin. Microbiol. 42(10): 4686-4696.
- **Doit**, C.; Loukil, C.; Geslin, P. and Bingen, E. (2002). Phenotypic and genetic diversity of invasive pneumococcal isolates recovered from French children. J Clin Microbiol 40: 2994-8.

- **Duarte**, R.S.; Barros, R.R.; Facklam, R.R. and Teixeira, L.M. (2005). Phenotypic and Genotypic Characteristics of Streptococcusporcinus Isolated from Human Sources. J. Clin. Microbiol. 43:4592–4601.
- Enright, M.C.; Spratt, B.G.; Kalia, A.; Cross, J.H. and Bessen, D.E. (2001). Multilocus sequence typing of Streptococcuspyogenes and their relationships between emm type and clone. Infect Immun; 69: 2416-27.
- Enright, M.C. and Spratt, B.G. (1998). A multilocus sequence typing scheme for Streptococcuspneumoniae: identification of clones associated with serious invasive disease. Microbiology 144, 3049-3060.
- Facklam, R.R. and Pigott, N. (1994). Description of phenotypic characteristics to aid in the identification of Streptococcuspneumoniae. In A. Totollan (ed.), Pathogenic streptococci: present and future. Lancer Publications, St. Petersburg, Russia.
- Falade, A.G.; Lagunju, I.A.; Bakare, R.A.; Odekanmi, A.A. and Adegbola, R.A. (2009). Invasive pneumococcal disease in children aged, 5 years admitted to 3 urban hospitals in Ibadan, Nigeria. Clin. Infect. Dis. 48:S190–196.
- **Fenoll**, A.; Munoz,R.; Garcia, E. and de la Campa,A.G. (1994). Molecular basis of the optochinsensitive phenotype of pneumococcus: characterization of the genes encoding the F0 complex of the Streptococcuspneumoniae and StreptococcusoralisH(+)-ATPases. Mol Microbiol 12(4): 587-98.
- **Fica**, A.; Fernande, J.; Ebensperger, G.; Cona, E.; Galanti, A. and Alonso, C. (2003). Molecular epidemiology of a Streptococcuspyogenes related nosocomial outbreak in a burn unit. Rev Med Chil; 131: 145-54.
- development and antibiotic susceptibility among Streptococcuspneumoniae isolates from cystic fibrosis samples and blood cultures. J Antimicrob Chemother 59: 301–304.
- Garcia-Medina, R.; Dunne, W.M.; Singh, P.K. and Brody, S.L. (2005). Pseudomonasaeruginosa acquires
- Fleih, M.T.; Al-Mathkhury, H.A. and Mahmod, Z. (2007). The pathological effect of peptidoglycan on rats'lungs part one: pathogenic bacteria Streptococcuspneumoniae. Journal of Al-Nahrain University. 10(2):87-93.
- Gonzalez-Rey, C.; Belin, A.M.; Jorbeck, H.; Norman, M.; Krovacek, K. and Henriques, B. (2003). RAPD-PCR and PFGE as tools in the investigation of an outbreak of beta-haemolytic Streptococcus group A in a Swedish hospital. Comp Immunol Microbiol Infect Dis; 26: 25-35.
- **Iwalokun**, B.A.; Fowora, M.; Akinloye, O.; Oluwadun, A.; Antonio, M. and Adegbola, R.A. (2012). Full Length Research paper A retrospective study of clinical Streptococcuspneumoniae isolates from four health facilities in South-West Nigeria. International Journal of Medicine and Medical Sciences Vol. 4(8):160-170.
- Kilian, M.; Poulsen, K.; Blomqvist, T.; Håvarstein, L.S.; Bek-Thomsen, M.; Tettelin, H. and Sørensen, U.B.S. (2008). Evolution of Streptococcuspneumoniae and Its Close Commensal Relatives. PLoS ONE 3(7): e2683.
- Obaro, S.; Lawson, L.; Essen, U.; Ibrahim, K.; Brooks, K.; Otuneye, A.; Shetima, D.; Ahmed, P.; Ajose, T.; Olugbile, M.; Idiong, D.; Ogundeji, D.; Ochigbo, C.; Olanipekun, G.; Khalife, W. and Adegbola, R. (2011). Community acquired bacteremia in young children from central Nigeria: A pilot study. BMC. Infect. Dis. 11:137.
- **Pettigrew**, M.M.; Fennie, K.P.; York, M.P.; Daniels, J. and Ghaffar, F. (2006). Variation in the presence of neuraminidase genes among Streptococcus pneumoniae isolates with identical sequence types. Infect. Immun. 74(6): 3360-3365.

- **Sambrook**, J. and Russell,R.W.(2001). Molecular cloning: A laboratory manual, 3rd ed. Cold spring harbor laboratory press, cold spring harbor, N.Y.
- Schlegel, L.; Grimont, F.; Grimont, P.A. and Bouvet, A. (2003). Identification of major Streptococcal species by rrn-amplified ribosomal DNA restriction analysis. J Clin Microbiol 41: 657-66.
- **Seppälä**, H.; He, Q.; Osterblad, M. and Huovinen, P.(1994). Typing of group A streptococci by random amplified polymorphic DNA analysis. J Clin Microbiol; 32: 1945-8.
- Shaaly, A.; Tellevik, M.G.; Langeland, N.; Hoiby, E.A. and Jureen, R. (2005). Comparison of serotyping, pulsed field gel electrophoresis and amplified fragment length polymorphism for typing of Streptococcuspneumoniae. J Med Microbiol 54: 467-72.
- **Shafeeq**, S.; Kuipers, O.P. and Kloosterman, T.G. (2013). Cellobiose-Mediated Gene Expression inStreptococcuspneumoniae: A Repressor Function of the Novel GntRType Regulator BguR. PLoS ONE 8(2): e57586.
- **Shakhnovich**, E.A.; King, S.J. and Weiser, J.N. (2002). Neuraminidase expressed by Streptococcuspneumoniae desialylates the lipopolysaccharide of Neisseria meningitides and Haemophilus influenza: a paradigm for interbacterial competition among pathogens of the human respiratory tract. Infect. Immun. 70: 7161-7164.
- **Sheppard**, C.L.; Harrison, T.G.; Morris, R.; Hogan, A. and George, R.C. (2004). Autolysin-targeted LightCycler assay including internal process control for detection of Streptococcuspneumoniae DNA in clinical samples. J Med Microbiol;53(Pt 3):189-95.
- **Shibl**, A.; Memish, Z. and Pelton, S. (2009). Epidemiology of invasive pneumococcal disease in the Arabian Peninsula and Egypt. Int J Antimicrob Agents 33(5): 410e1-9.
- Shivshankar, P.; Sanchez, C.; Rose, L.F. and Orihuela, C.J. (2009). The Streptococcuspneumoniae adhesin PsrP binds to Keratin 10 on lung cells. Mol Microbiol 73: 663–679.
- **Shortridge**, V.D.; Doern, G.B.; Brueggemann, A.B.; Beyer, J.M. and Flamm, R.K. (1999). Prevalence of macrolide resistance mechanisms in Streptococcuspneumoniae isolates from a multicenter antibiotic resistance surveillance study conducted in the United States in 1994–1995. Clinical Infectious Diseases 29:1186-1188.
- **Shundi**, L.; Surdeanu, M. and Damian, M. (2000). Comparison of serotyping, ribotyping and PFGE for distinguishing group A Streptococcus strains isolated in Albania. Eur J Epidemiol; 16: 257-63.
- Shutt, C.K.; Samore, M. and Carroll, S.K.C. (2004). Comparison of the Denka Seiken Slide Agglutination Method to the Quellung Test for Serogrouping of Streptococcuspneumoniae Isolates. J. Clin. Microbiol, 42(3):1274-1276.
- **Siberry**, G.; Brahmadathan, K.N.; Pandian, R.; Lalitha, M.K.; Steinhoff, M.C. and John, T.J. (2001). Comparison of different culture media and storage temperatures for the long-term preservation of Streptococcus pneumoniae in the tropics. Bull. W. H. O. 79:43–47.
- **Steinberg**, J.J. ;Levine,D.S. ; Desiderio,D. and Hanna,B.A.(1988). Serotypes of Streptococcuspneumoniae recovered from a large urban hospital population: the Bellevue experience from 1973-1984. Lab. Med. 19:741-743.
- **Steinfort**, C.; Wilson, R.; Mitchell, T.; Feldman, C.; Rutman, A.; Todd, H.; Sykes, D.; Walker, J.; Saunders, K.; Andrew, P.W. and et al. (1989). Effect of Streptococcuspneumoniae on human respiratory epithelium in vitro. Infect Immun 57(7): 2006–13.

- Suzuki, N.; Yuyama, M.; Maeda, S.; Ogawa, H.; Mashiko, K. and Kiyoura, Y. (2006). Genotypic identification of presumptive Streptococcuspneumoniae by PCR using four genes highly specific for S. pneumoniae. J Med Microbiol;55(Pt 6):709-14.
- **Tonnaer**, E.L.; Rijkers, G.T.; Meis, J.M.; Klaassen, C.H.; Bogaert, D.; Hermans, P.W. and Curfs, J.H. (2005). Genetic Relatedness between Pneumococcal Populations Originating from the Nasopharynx, Adenoid, and Tympanic Cavity of Children with Otitis Media. Journal of Clinical Microbiology, 43(7):3140–3144.